# Evaluation of *de novo*Transcriptome Assemblies from RNA-Seq Data

with Bo Li and Colin Dewey CIBM Seminar, February 12, 2012

#### Background - The Transcriptome

Definition: The collection of RNA molecules in a cell or sample.

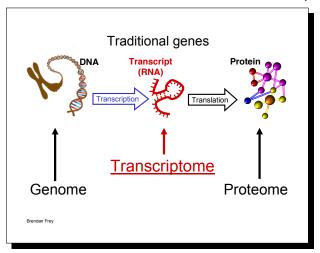
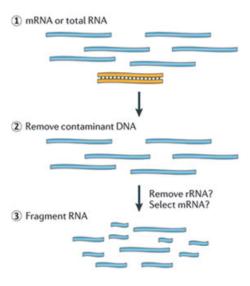
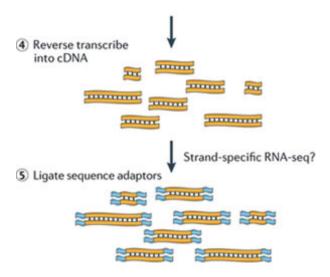


Figure from Brendan Frey, http://www.psi.toronto.edu/isit2006/.

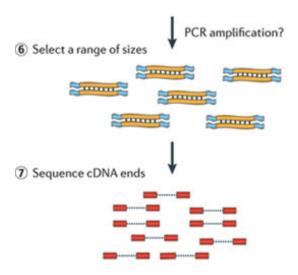
# Background - RNA-Seq



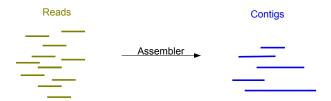
## Background - RNA-Seq



# Background - RNA-Seq

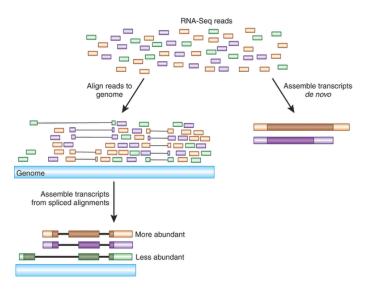


# Background - Assembly



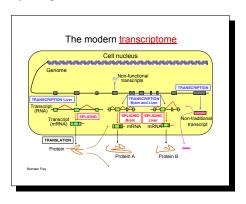
A contig is a continguous subsequence of a transcript sequence.

## Background - Assembly

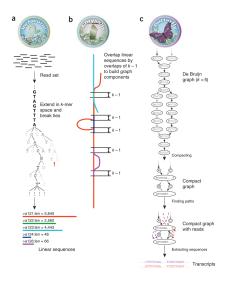


## **Background - Complications**

- Non-uniform expression.
- Alternative splicing.



## Background - de novo Assembly



#### **Evaluation Problem**

Without the ground truth reference transcript set,
determine which assembly is best
based only on the RNA-Seq data
from which the assemblies were constructed.

#### **Evaluation Problem - Desiderata**

- Start from first principles.
- Avoid trivialalities.
- Achieve the same ordering as a simple reference-based score.

Non-solution: N50, the largest n such that the contigs with length  $\geq n$  compose at least 50% of the total bases of the contigs set.

#### **Our Contributions**

- A score that satisfies the given desiderata.
- A reference-based precision/recall framework for transcriptome assembly.
- ▶ A software package, DETONATE, that implements the above.
- ► A comprehensive meta-evaluation of the score.

# The Score Is Based on a Probability Model

Our score:  $P(assembly, reads) \propto P(assembly|reads)$ .

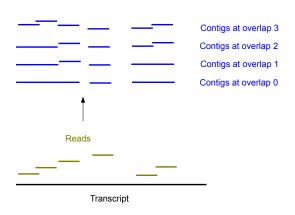
$$P(\text{assembly}, \text{reads})$$

$$= \int P(\text{assembly}, \text{coverage}, \text{reads}) \, d\text{coverage}$$

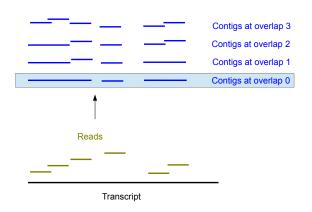
$$= \int \underbrace{P(\text{assembly}, \text{coverage})}_{\text{prior}} \underbrace{P(\text{reads}|\text{assembly}, \text{coverage})}_{\text{likelihood}} \, d\text{coverage}$$

A contig's "coverage" is the expected number of reads generated from each position of the contig's original transcript.

# The Probability Model Is Based on Ideal Assembly



# The Probability Model Is Based on Ideal Assembly



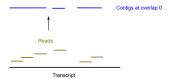
## The Probability Model - Prior

```
P(\text{assembly}, \text{reads}) \\ = \int P(\text{assembly}, \text{coverage}, \text{reads}) \, d\text{coverage} \\ = \int \underbrace{P(\text{assembly}, \text{coverage})}_{\text{prior}} \underbrace{P(\text{reads}|\text{assembly}, \text{coverage})}_{\text{likelihood}} \, d\text{coverage}
```

#### The Probability Model - Prior

#### Generative story:

- ▶ Transcript lengths  $\sim_{iid}$  negative binomial.
- Given the transcript lengths:
  - ▶ Transcript sequences  $\sim_{iid}$  uniform.
  - Number of reads starting at each position of a transcript
     Poisson (mean = coverage).
- The assembly is formed from the reads at overlap 0.



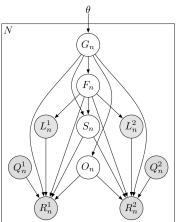
One can work out a recurrence for the prior probability of the assembly and coverage.

# The Probability Model - Likelihood

$$P(\text{assembly}, \text{reads}) \\ = \int P(\text{assembly}, \text{coverage}, \text{reads}) \, d\text{coverage} \\ = \int \underbrace{P(\text{assembly}, \text{coverage})}_{\text{prior}} \underbrace{P(\text{reads}|\text{assembly}, \text{coverage})}_{\text{likelihood}} \, d\text{coverage}$$

## The Probability Model - Likelihood

Previous work, RSEM, introduced a generative model of reads, given transcripts and their expression.



## The Probability Model - Likelihood

#### Key observation:

► Generating from contigs = generating from transcripts, except that contigs are guaranteed to be covered by reads.

#### Therefore, we stipulate:

```
\begin{split} &P(\text{reads}|\text{assembly}, \text{coverage}) \\ &= \frac{P_{RSEM}(\text{reads}|\text{transcripts} = \text{assembly}, \text{expression} = f(\text{coverage}))}{P_{RSEM}(\text{reads cover assembly}|\text{transcripts} = \text{assembly}, \text{expression} = f(\text{coverage}))} \end{split}
```

## The Probability Model - Marginalization

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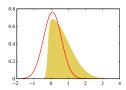
Approximate the integral by BIC:

log P(assembly, reads)

$$=\log\int P(\text{assembly}, \text{coverage}, \text{reads}) d\text{coverage}$$

= log 
$$P$$
(assembly, reads|coverage\*)  $-\frac{1}{2}M\log N$ 

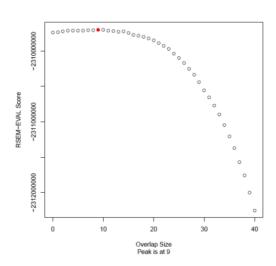
where M = number of contigs, N = number of reads, coverage\* = maximum likelihood estimate.



## Experiment 0 - Setup

Goal: Make sure we have avoided trivialities.

- Procedure:
  - Construct ideal assembly at every possible overlap.
  - Compute score.
- Desired result: Best overlap is fairly close to 0.

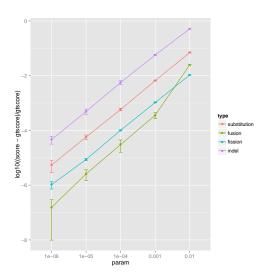


#### Experiment 1 - Setup

Goal: Make sure the true best assembly has the best score, on average.

#### Procedure:

- Construct ideal assembly at overlap 0.
- Perturb this assembly:
  - Substitution substitute a base.
  - Fusion join two contigs into one contig.
  - Fission split one contig into two contigs.
  - Indel insert or delete a fragment from a contig.
- Compute score for ideal and perturbed assemblies.
- ▶ Desired result: The ideal assembly has the best score.

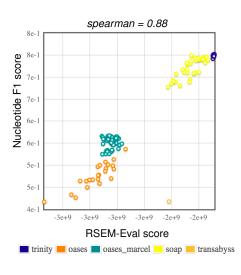


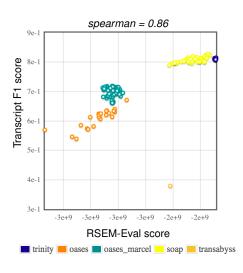
#### Experiment 2 - Setup

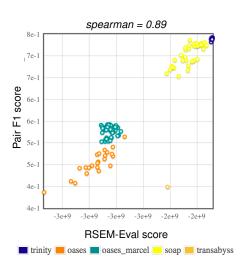
- Goal: Study the correlation between our score and simple reference-based scores.
- Five datasets:
  - Mouse from Trinity paper.
  - Mouse from Oases paper.
  - Yeast from Trinity paper.
  - Axolotl from Thompson lab.
  - Simulated mouse.
- ► ~100 assemblies per dataset, using:
  - Trinity.
  - Oases.
  - SOAPdenovo-trans.
  - Trans-ABySS.

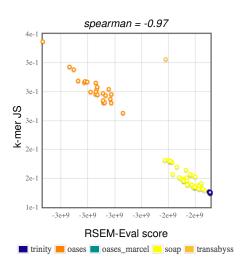
#### Experiment 2 - Setup

- 3 reference-based F1 scores (harmonic mean of precision and recall):
  - Nucleotide F1.
  - Transcript F1.
  - Pair F1.
- 1 reference-based "k-mer" score:
  - Jensen-Shannon divergence between k-mer distributions.
- Procedure:
  - For each assembly: compute our de novo score and each reference-based score.
- Expected result:
  - Monotone relationship between the scores.









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