

# Evaluation of *de novo* transcriptome assemblies from RNA-Seq data

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based on work with Bo Li, Yongsheng Bai, Mike Collins,  
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# De novo transcriptome assembly

```
read 1:  AGCATCGCGT
read 2:  CGTTGCGTCC
read 3:  CGTCCCGCGC
read 4:  GCGCGCTTAG
read 5:  GCTACTCTCA
read 6:  TACTCTCACA
```

Reads



```
AGCATCGCGT
  CGTTGCGTCC
    CGTCCCGCGC
      GCGCGCTTAG

GCTACTCTCA
  TACTCTCACA
```



```
contig 1: AGCATCGCGTTGCGTCCCGCGCGCTTAG
contig 2:  GCTACTCTCACA
```

*De novo* assembly

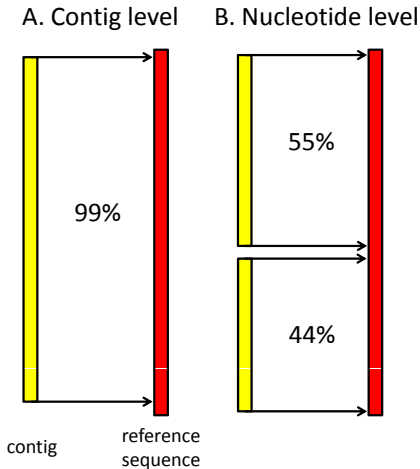


## Two classes of assembly evaluation measures

- ▶ Reference-based: compare assembly to ground truth reference.
- ▶ Reference-free: evaluate assembly without reference.



# Reference-based evaluation



Recall values

	Contig	Nucleotide
A	100%	99%
B	0%	99%



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- ▶ Statistical model-based scores for evaluating genome (CGAL: Rahman and Pachter, 2013) and metagenome (Genovo: Laserson et al., 2011; ALE: Clark et al., 2013) assemblies.



Our contribution is a **reference-free** transcriptome assembly **scoring function**, which can be used to choose the best assembly from a collection of candidate *de novo* assemblies when no ground-truth reference is available. The score is **based on a statistical model** of the process of RNA-Seq read generation and of “true” transcriptome assembly.



$$\begin{aligned}\text{score}(\text{assembly}, \text{reads}) &= \log P(\text{assembly}, \text{reads}) \\ &= \log \int P(\text{assembly}|\lambda)P(\text{reads}|\text{assembly}, \lambda) dP(\lambda) \\ &\approx \underbrace{\log P(\text{assembly}|\lambda^*)}_{\text{prior}} + \underbrace{\log P(\text{reads}|\text{assembly}, \lambda^*)}_{\text{likelihood}} \\ &\quad - \underbrace{\frac{1}{2}N_{\text{contigs}} \log N_{\text{reads}}}_{\text{BIC penalty}}\end{aligned}$$

A contig's “coverage”  $\lambda_i$  is the expected number of reads generated from each position of the contig's parent transcript, and  $\lambda^*$  is the maximum likelihood estimate.





The prior distribution over assemblies is specified indirectly:

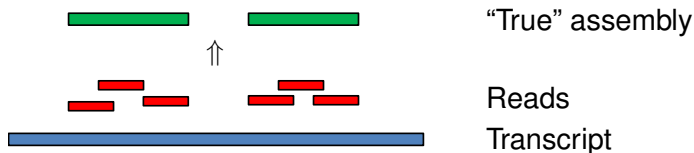
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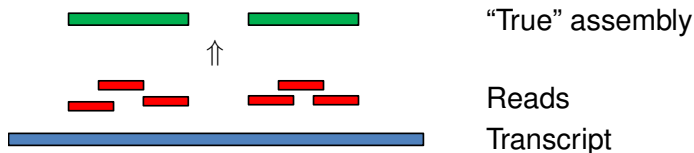
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- ▶ The above induces a distribution over assemblies.



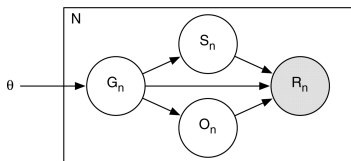
Practical contribution of the prior:

- ▶ Penalizes assemblies whose contigs have aberrant lengths relative to the coverage.
- ▶ Penalizes assemblies with too many nucleotides.



# The data likelihood $P(\text{reads}|\text{assembly}, \lambda)$

RSEM (Li et al., 2010), introduced a generative model of reads, given transcripts and their expression:



where

- ▶  $\theta_j$  is the expression of transcript  $j$ .
- ▶  $N$  is the number of reads.
- ▶  $G_n$  is the transcript read  $n$  comes from.
- ▶  $S_n$  is the start position of read  $n$  within its transcript.
- ▶  $O_n$  is the orientation of read  $n$  within its transcript.
- ▶  $R_n$  is read  $n$ .



Key observation:

- ▶ Generating from contigs  $\equiv$  generating from transcripts, except that contigs are guaranteed to be covered by reads.



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Therefore, we define the likelihood to be the probability of the reads given the contigs, according to RSEM's model, divided by the probability that the contigs are covered by reads.

Practical contribution of the likelihood:

- ▶ On one hand, the likelihood penalizes contigs that are not well-supported by reads.
- ▶ On the other hand, the likelihood penalizes assemblies that do not make use of all the reads.





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- ▶ Simulate RNA-Seq data.
- ▶ Construct the “true” assembly.
- ▶ Perturb this assembly:
  - ▶ Substitution - substitute a base.
  - ▶ Fusion - join two contigs into one contig.
  - ▶ Fission - split one contig into two contigs.
  - ▶ Indel - insert or delete a fragment from a contig.



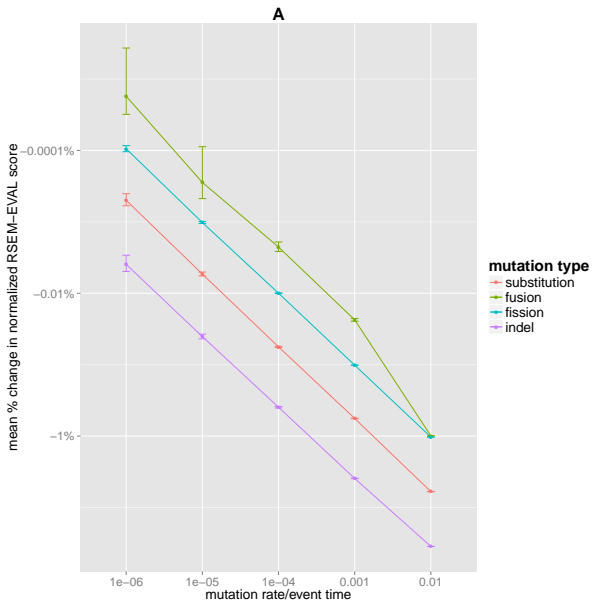
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- ▶ Compute score for “true” and perturbed assemblies.



# Experiment 1 - Random perturbation - Results



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Procedure:

- ▶ For each dataset (real mouse and simulated mouse):
  - ▶ Create  $\sim 200$  assemblies, by running several assemblers with different parameter settings.
  - ▶ For each assembly, compute:
    - ▶ Our model-based score.
    - ▶ Contig and nucleotide F1.
    - ▶ Our reference-based  $k$ -mer compression score (next slide).

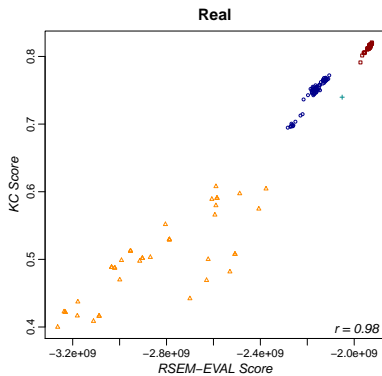
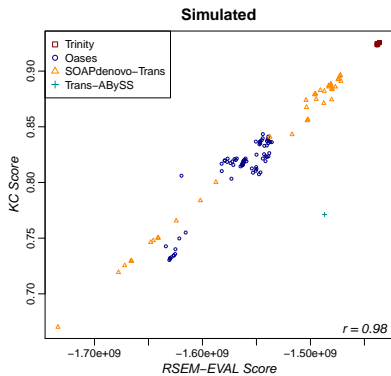


$k$ -mer compression (KC) score = weighted  $k$ -mer recall (WKR)  
– inverse compression ratio (ICR).

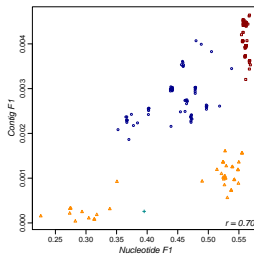
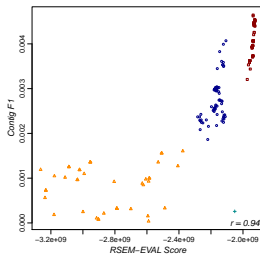
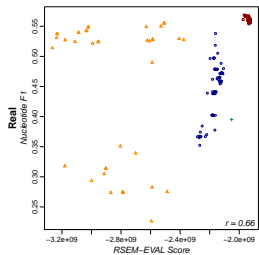
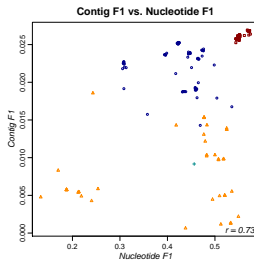
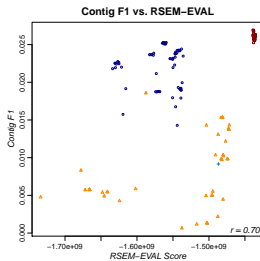
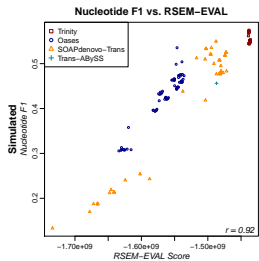
- ▶ WKR = assembly's recall of the  $k$ -mers present in the reference sequences, with each  $k$ -mer weighted by its relative frequency within the reference transcriptome.
- ▶  $ICR = \frac{\text{number of bases in the assembly}}{\text{number of bases in the set of reads}}$ .



# Experiment 2 - Correlation - Results



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# Thanks

## Software:

- ▶ DETONATE: <http://deweylab.biostat.wisc.edu/detonate/>



# Extra slide - RSEM-EVAL Example

RNA-Seq  
3,818 reads

Rpl24

Trinity

Oases

SOAPdenovo-Trans

Score Assembly	Likelihood score	Prior score	BIC score	RSEM-EVAL score
	-86542	-876	-8	-87426
	-198666	-2771	-29	-201466
	-254310	-613	-12	-254935



# Extra slide - Runtime comparison

Program	Assembly T		Assembly O		Assembly S	
	Runtime	Memory	Runtime	Memory	Runtime	Memory
RSEM-EVAL*	1h 4m 57s	2.02 GB	4h 40m 36s	8.18 GB	34m 57s	1.23 GB
Genovo	6d 11h 54m 3s	192.23 GB	> 1 week	-	4d 15h 3m 3s	188.79 GB
ALE*	12h 39m 36s	0.67 GB	6d 23h 23m 13s	2.31 GB	7h 33m 1s	0.59 GB
REF-EVAL, contig**	3s	0.19 GB	8s	0.33 GB	2s	0.2 GB
REF-EVAL, nucleotide**	8s	0.39 GB	33s	1.27 GB	6s	0.33 GB
REF-EVAL, KC score	1m 18s	2.09 GB	1m 30s	2.37 GB	1m 13s	2.03 GB
Bowtie	15m 42s	0.11 GB	1h 1m 38s	0.31 GB	11m 16s	0.1 GB
Blat	35m 14s	0.0 GB	1h 51m 1s	0.01 GB	28m 19s	0.0 GB

\* Plus time to run Bowtie. We calculate Bowtie statistics separately because ALE takes Bowtie alignments as input.

\*\* Plus time to run Blat.



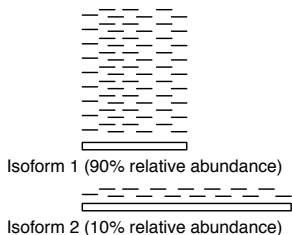


	KC Score	Contig F1	Nucleotide F1
RSEM-EVAL Score	0.99	0.83	0.46
Genovo Score	0.96	0.80	0.53
ALE Score	0.64	0.45	0.62
N50	0.22	0.33	-0.31
Number of Nucleotides in Assembly	0.13	0.29	-0.21
Number of Unique Proteins Matched	0.68	0.81	0.73
Average Ortholog Hit Ratio	0.31	0.31	-0.19

**Table 1** The Spearman rank correlation coefficient of the scores assigned by several alternative transcriptome assembly evaluation measures, described in the main text, to the reference-based scores from REF-EVAL. The evaluated assemblies were produced by Trinity, Oases, SOAPdenovo-Trans, and Trans-ABYSS, based on the subset of reads in the real (strand non-specific) mouse data that align to genes on chromosome 1. This subset was used in the interest of computational efficiency of the alternative measures.



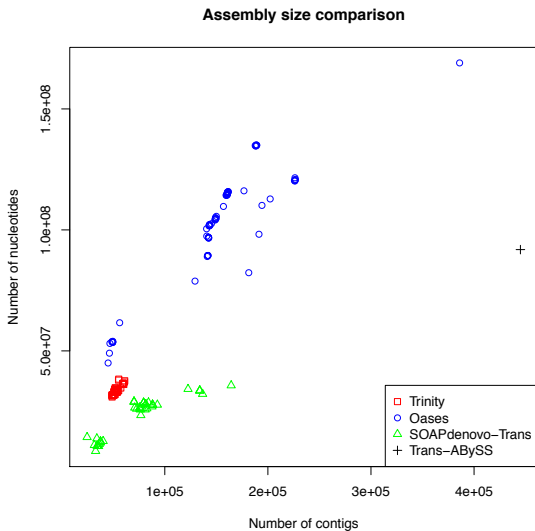
## Extra slide - Comparison on an example



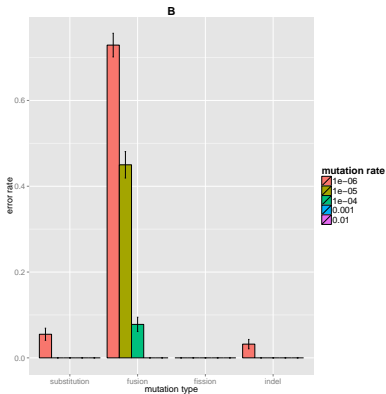
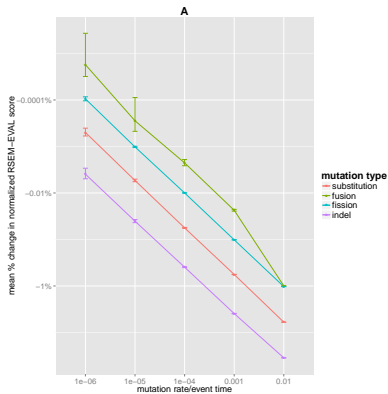
<b>Score</b> <b>Assembly</b>	<b>RSEM-EVAL</b>	<b>GENOVO</b>	<b>ALE</b>
<b>Truth</b>	-43720	-19557	-116316
<b>Long only</b>	-44403	-18199	-88905
<b>Short only</b>	-104963	-68997	-52090



# Extra slide - Different parameters give very different assemblies



# Extra slide - Experiment 1 - Random perturbation - Results



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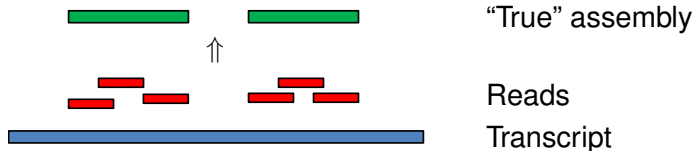
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- ▶ Based on the above, one can work out a recurrence for the prior probability of the assembly.

